



Evaluation of Simpler Criteria for Diagnosing Allergic Bronchopulmonary Aspergillosis Complicating Asthma

Ritesh Agarwal^{1*†}, Puneet Saxena^{1†}, Valliappan Muthu¹, Inderpaul Singh Sehgal¹, Sahajal Dhooria¹, Kuruswamy Thurai Prasad¹, Ashutosh Nath Aggarwal¹ and Arunaloke Chakrabarti²

¹ Department of Pulmonary Medicine, Postgraduate Institute of Medical Education and Research (PGIMER), Chandigarh, India, ² Department of Medical Microbiology, Postgraduate Institute of Medical Education and Research (PGIMER),

OPEN ACCESS

Edited by:

Chandigarh, India

Neil Andrew Robert Gow, University of Exeter, United Kingdom

Reviewed by:

Stéphane Ranque, Aix-Marseille Université, France Vishukumar Aimanianda, Institut Pasteur, France

*Correspondence:

Ritesh Agarwal agarwal.ritesh@outlook.in

[†]These authors have contributed equally to this work and share first authorship

Specialty section:

This article was submitted to Fungal Pathogenesis, a section of the journal Frontiers in Cellular and Infection Microbiology

Received: 25 January 2022 Accepted: 07 March 2022 Published: 25 March 2022

Citation:

Agarwal R, Saxena P, Muthu V, Sehgal IS, Dhooria S, Prasad KT, Aggarwal AN and Chakrabarti A (2022) Evaluation of Simpler Criteria for Diagnosing Allergic Bronchopulmonary Aspergillosis Complicating Asthma. Front. Cell. Infect. Microbiol. 12:861866. doi: 10.3389/fcimb.2022.861866 **Background:** The modified International Society for Human and Animal Mycology (ISHAM) allergic bronchopulmonary aspergillosis (ABPA) working group (AWG) criteria lists up to five components for diagnosing ABPA in asthmatics. Whether eliminating specific components of the existing criteria would have the same diagnostic utility as the original remains unknown.

Objective: To evaluate the performance of several simplified criteria for diagnosing ABPA.

Methods: We compared the performance of seven new criteria (after excluding or modifying one or more of the components of the ISHAM-AWG criteria) with the modified ISHAM-AWG criteria in asthmatic subjects using latent class analysis (LCA). We also tested the performance of the newer criteria using accuracy measures against a multidisciplinary team (MDT) diagnosis of ABPA. We considered the diagnostic accuracy of the newer criteria to be acceptable if the correct classification and false-negative rates were >95% and <5%, respectively, on an MDT evaluation.

Results: We analyzed data from 543 asthmatic subjects (58.8% women; mean age, 36.8 years). Using LCA, the sensitivity of the *A.fumigatus*-specific IgE-based criteria ranged from 92-99%, while the specificity varied between 92% and 100%. The MDT diagnosed ABPA in 106 (19.5%) subjects. Using MDT as the reference standard, the correct classification and false-negative rates were >95% for three of the seven and <5% for four of the seven newer criteria.

Conclusions: We found several of the newly developed criteria to perform, like the modified ISHAM-AWG criteria, for diagnosing ABPA complicating asthma. A prospective study in current clinical algorithms is required for validating our observations.

Keywords: allergic bronchopulmonary mycosis (ABPM), *Aspergillus*, fungal sensitization, allergy, latent class analysis (LCA), bronchial asthma, cystic fibrosis

1

INTRODUCTION

Allergic bronchopulmonary aspergillosis (ABPA) is a pulmonary disorder caused by complex hypersensitivity reactions targeted against *Aspergillus fumigatus* colonizing the airways of patients with asthma (and cystic fibrosis) (Agarwal et al., 2022). ABPA patients typically present with difficult-to-control asthma, recurrent pulmonary opacities, and bronchiectasis (Kosmidis and Denning, 2015). Several criteria based on a composite of imaging and immunological parameters are available for diagnosing ABPA (Rosenberg et al., 1977; Agarwal et al., 2010; Agarwal et al., 2013; Agarwal et al., 2013; Dhooria and Agarwal, 2014; Asano et al., 2021; Saxena et al., 2021). Rosenberg et al. were the first to suggest a criterion for diagnosing ABPA (Rosenberg et al., 1977). Subsequently, several modifications to these criteria have been proposed to incorporate the advances in immunological tests.

In 2013, the ABPA working group (AWG) of the International Society for Human and Animal Mycology (ISHAM) had framed a criterion for diagnosing ABPA in asthmatics (Agarwal et al., 2013). The ISHAM-AWG criterion relies on certain thresholds of a combination of several immunological and other tests. Although this criterion is a refinement over previously available diagnostic algorithms, it would be interesting to both clinicians and patients if the diagnostic standards could be made even easier. Recently, we proposed specific changes to the ISHAM-AWG criteria that made the criteria simpler and resulted in better sensitivity and specificity (Saxena et al., 2021). The current study aimed to evaluate the performance of several simpler criteria for diagnosing ABPA.

METHODS

We conducted a retrospective analysis of prospectively collected data (July 2017 and September 2018) at our institute's chest clinic. The Institute Ethics Committee approved the study protocol. We obtained written informed consent from all the participants.

Patients

We have published the detailed methodology and the patient details previously (Saxena et al., 2021). Briefly, we included consecutive cases of physician-diagnosed bronchial asthma. We classified the asthma severity based on the 2004 Global Initiative for Asthma (GINA) recommendations that adjusts for the effect of treatment on disease severity (Global Initiative for Asthma (GINA), 2004).

Study Procedure

We performed spirometry, *Aspergillus* skin test (AST), *A.fumigatus*-specific IgE and IgG, serum total IgE, peripheral blood total eosinophil count (TEC), chest radiograph, and computed tomography (CT) of the chest in all the asthmatic subjects. We performed AST intradermally using the crude *A.fumigatus* antigen (Alcit India, New Delhi, India), and an immediate cutaneous hyperreactivity was considered a positive AST (Agarwal et al., 2013). Serum total IgE, *A.fumigatus*-specific IgE, and IgG were assayed using the commercially available fluorescent enzyme immunoassay (Phadia 100, Thermofisher Scientific, Uppsala, Sweden) (Agarwal et al., 2016; Agarwal et al., 2017). We classified the chest radiographic findings as transient or permanent (Agarwal et al., 2016). We obtained CT of the thorax using a multidetector CT scanner and defined bronchiectasis according to previously described criteria (Agarwal et al., 2011). We performed spirometry on an ultrasonic flow-sensing spirometer and interpreted the spirometric values, namely the forced expiratory volume in the first second (FEV1) and the forced vital capacity (FVC), using the Indian guidelines (Aggarwal et al., 2019).

Newer, Simpler Criteria

The modified ISHAM-AWG criterion requires up to 5 components for diagnosing ABPA in asthmatics (**Table 1**). We tested five criteria (criteria 1, 2, 5, 6, and 7) with only four components (eliminating TEC or *A.fumigatus*-specific IgG or CT) and two criteria (criteria 3 and 4) with only three components (eliminating both TEC and *A.fumigatus*-specific IgG). All the criteria included *A.fumigatus*-specific IgE except criteria 4 that replaced it with AST. Criteria 1 and 2 were like criteria 5 and 6, respectively, except for a higher threshold for *A.fumigatus*-specific IgE (0.5 kUA/L instead of 0.35 kUA/L). Criteria 4 was similar to criteria 3, except it replaced *A.fumigatus*-specific IgE with AST. Finally, criteria 7 was solely an immunological criterion, and we excluded CT chest.

TABLE 1 | Various proposed simpler criteria for allergic bronchopulmonary aspergillosis (ABPA) evaluated in the current study.

Modified ISHAM-AWG criteria

Presence of both: (1) serum Af-specific IgE >0.35 kUA/L, and (2) serum total IgE >500 IU/mL AND two of the following: (1) serum Af-specific IgG >27 mgA/L; (2) bronchiectasis on CT chest; (3) TEC >500 cells/ μ L

New criteria

Criteria 1: Both: (1) serum Af-specific IgE >0.5 kUA/L, and (2) serum total IgE >500 IU/mL AND any of the following: (1) serum Af-specific IgG >27 mgA/L; (2) bronchiectasis on CT chest

Criteria 2: Both: (1) serum Af-specific IgE >0.5 kUA/L, and (2) serum total IgE >500 IU/mL AND any of the following: (1) bronchiectasis on CT chest; (2) TEC >500 cells/µL

Criteria 3: All the following: (1) serum Af-specific IgE >0.35 kUA/L; (2) serum total IgE >500 IU/mL; (3) bronchiectasis on CT chest

Criteria 4: All the following: (1) type 1 Aspergillus skin test positive; (2) serum total IgE >500 IU/mL; (3) bronchiectasis on CT chest

Criteria 5: Both: (1) serum Af-specific IgE >0.35 kUA/L, and (2) serum total IgE >500 IU/mL AND any of the following: (1) serum Af-specific IgG >27 mgA/L; (2) bronchiectasis on CT chest

Criteria 6: Both: (1) serum Af-specific IgE >0.35 kUA/L, and (2) serum total IgE >500 IU/mL AND any of the following: (1) bronchiectasis on CT chest; (2) TEC >500 cells/ μ L

Criteria 7: Both: (1) serum Af-specific IgE >0.35 kUA/L, and (2) serum total IgE >500 IU/mL AND any of the following: (1) serum Af-specific IgG >27 mgA/L; (2) TEC >500 cells/ μ L

Af, Aspergillus fumigatus; AWG, ABPA working group; CT, computed tomography; ISHAM, international society for human and animal mycology; TEC, peripheral blood total eosinophil count.

Objectives

The study objectives were to (1) compare the diagnostic performance of seven simpler standards (**Table 1**) with the recently published modified ISHAM-AWG criteria for diagnosing ABPA, using latent class analysis (LCA); (2) evaluate the diagnostic performance of the different newer benchmarks for detecting ABPA against a multidisciplinary team (MDT) diagnosis of ABPA.

Statistical Analysis

We analyzed data using the commercial statistical package SPSS version 22.0 (IBM SPSS Inc., Armonk NY, US) and present the data as number (percentage) or mean (standard deviation [SD]). We performed LCA using the R version of the TAGS software (Pouillot et al., 2002; Agarwal et al., 2013; Saxena et al., 2021). For performing LCA, we divided the asthma population into mildto-moderate (consisting of mild intermittent, mild persistent, and moderate persistent asthma) or severe (comprising of severe persistent asthma), according to the 2004 GINA guidelines (Global Initiative for Asthma (GINA), 2004). We also analyzed the difference in LCA results when the study population is not divided into two groups (single group). We calculated the sensitivity and specificity (with corresponding 2.5-97.5% bootstrap confidence intervals [CI]), obtained by bootstrapping 5000 samples) using the maximum likelihood (ML) estimates. We used the Newton-Raphson and the expectationmaximization algorithms for deriving the ML estimates (Pouillot et al., 2002). We assumed conditional independence using the goodness-of-fit test and subsequently deduced the residual correlations between tests. A p-value >0.05 and the random distribution of the residuals near-zero indicated a good fit of the model. We used the DAG_stat application for calculating the sensitivity, specificity, correct classification rate (efficiency), and false-negative rate with 95% CI of the various criteria using MDT diagnosis as the reference standard (Mackinnon, 2000). We assumed the diagnostic accuracy of the newer criteria to be acceptable if the efficiency was >95% and the false-negative rate <5%.

RESULTS

We included 543 asthmatic subjects (58.8% women) with a mean (SD) age of 36.8 (13.9) years. There were 338 and 205 subjects with mild-to-moderate and severe asthma, respectively. The proportion of subjects with aspergillus skin test positivity, serum total IgE level >500 IU/mL, serum *A.fumigatus*-specific IgE values >0.35 and >0.5 kUA/L, serum *A.fumigatus*-specific IgG >27 mgA/L, and peripheral blood TEC >500 cells/µL were similar between the two groups (**Table 2**). We encountered bronchiectasis in 24.3% of the study subjects.

Objective 1

We compared seven newer criteria with the modified ISHAM-AWG criteria using LCA (Table 3). The sensitivity of the A.fumigatus-specific IgE-based criteria (criteria 1, 2, 3, 5, 6, and 7) ranged from 92-99%, while the specificity varied between 92% and 100%. The diagnostic performance of A.fumigatus-specific IgE dependent criteria was almost similar to the modified ISHAM-AWG criteria. The sensitivity was lowest for the new criteria 4 [also known as minimal diagnostic criteria for ABPA (Schwartz and Greenberger, 1991)]; however, the specificity was nearly 100%. When the skin test in the minimal essential criteria was replaced with A.fumigatus-specific IgE (Table 3, criteria 3), the sensitivity of the minimal criteria increased to 92% with no loss of specificity. The specificity was lowest for the new criteria 7 that excluded imaging, although the sensitivity was 93.5%. The ML estimate for the model was -857.8 (deviance, 631.8). The p-value for the goodness-of-fit test for conditional independence was 0.00001, indicating that the model does not fit. We found the results similar even if the population is kept as a single group (Table 3).

TABLE 2 | Select baseline characteristics of the study population. Total (n=543) Mild-to-moderate asthma (n=338) Severe asthma (n=205) P value 36.8 ± 13.9 34.9 ± 13.5 39.9 ± 13.8 0.0001 Age (vears) 319 (58.8) 186 (55.0) 133 (64.9) 0.024 Female sex 236 + 52235 + 52238 + 520.55 Body mass index (Kg/m²) Asthma duration (years) 10.2 ± 9.5 9.0 ± 8.8 12.0 ± 10.3 0.0001 Spirometry FEV1 (% predicted) 735 + 256777 + 240666 + 2670.0001 FVC (% predicted) 82.7 ± 20.7 85.9 ± 19.8 77.2 ± 21.0 0.0001 Immunologic investigations 0.86 Type 1 Af skin test positive 164 (30.2) 103 (30.5) 61 (29.8) Serum Af-specific IgE >0.35 kUA/L 219 (40.5) 133 (39.5) 86 (42.2) 0.54 Serum Af-specific IgE >0.5 kUA/L 201 (37.0) 123 (36.4) 78 (33.0) 0.70 Serum Af-specific IgG >27 mgA/L 194 (35.7) 119 (35.2) 75 (36.6) 0.75 Peripheral blood eosinophil count >500 cells/µL 223 (41.1) 132 (39.1) 91 (44.4) 0.22 Serum total IgE level >500 IU/mL 300 (55.3) 186 (55) 114 (55.6) 0.90 **Bronchiectasis on CT chest** 59 (28.8) 0.059 132 (24.3) 73 (21.6)

All values are represented as mean ± SD or number (percentage) unless otherwise indicated.

Af, Aspergillus furnigatus; CT, computed tomography; FEV1, forced expiratory volume in the first second; FVC, forced vital capacity.

Mild-to-moderate asthma consisted of mild intermittent, mild persistent, and moderate persistent, while severe asthma comprised of severe persistent category as per the 2004 GINA quidelines

	Тwo рој	oulations	Single population		
	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)	
Modified ISHAM-AWG criteria	94.4 (89.7-98.3)	100 (100-100)	94.4 (89.7-98.3)	100 (100-100)	
New criteria					
Criteria 1	96.3 (92.4-99.2)	97.7 (96.2-99.1)	93.5 (88.6-97.9)	97.7 (96.2-99.1)	
Criteria 2	98.1 (95.2-100)	95.4 (93.3-97.3)	98.1 (95.0-100)	95.4 (93.4-97.3)	
Criteria 3	91.7 (86.0-96.6)	100 (100-100)	91.6 (86.0-96.4)	100 (100-100)	
Criteria 4	73.1 (64.4-81.3)	99.8 (99.3-100)	73.1 (64.7-81.3)	99.8 (99.3-100)	
Criteria 5	99.1 (96.9-100)	97.7 (96.2-99.1)	99.1 (96.7-100)	97.7 (96.2-99.1)	
Criteria 6	99.1 (96.9-100)	94.0 (91.7-96.1)	99.1 (96.7-100)	94.0 (91.7-96.2)	
Criteria 7	93.5 (88.6-97.9)	91.7 (88.9-94.2)	93.5 (88.5-97.9)	91.7 (89.1-94.3)	

The values in parentheses are confidence intervals that correspond to 2.5-97.5% bootstrap confidence intervals (CI) obtained by bootstrapping 5000 samples

ISHAM-AWG, international society for human and animal mycology ABPA working group criteria

The p-value for goodness-of-fit for the models is <0.05, indicating that the model does not fit.

The details of the various criteria are provided in Table 1.

Objective 2

We further compared the diagnostic performance of various criteria using an MDT diagnosis as the reference standard. The MDT adjudicated 106 cases to have ABPA (**Table 4**). The correct classification rate was >95% for criteria 1, 3, and 5 (**Table 4**). The false-negative rate was <5% for all criteria except for criteria 3 (11.3%), 4 (26.5%), and 7 (26.3%).

DISCUSSION

We found several simpler criteria to perform reasonably well than the modified ISHAM-AWG criteria. An optimal diagnostic criterion should be valid, reliable, and straightforward. Moreover, the criterion should have a clearly defined diagnostic threshold. The modified ISHAM-AWG meets most of these attributes but involves several tests, and less complex criteria would be welcome. While minimal criteria have been proposed in the past (Schwartz and Greenberger, 1991), none have been evaluated for their performance. We designed seven criteria and compared them with the modified ISHAM-AWG criteria. We found no benefit on LCA and MDT evaluation by increasing the *A.fumigatus*-specific IgE threshold from 0.35 to 0.5 kUA/L (criteria 1 and 2 vs. 5 and 6). The minimal essential diagnostic criteria for ABPA (Aspergillus skin test positivity, raised serum total IgE, and bronchiectasis; criteria 4) (Schwartz and Greenberger, 1991) had a sensitivity of only about 74%, thus potentially missing many cases despite having a nearly 100% specificity. There was a considerable increase in the sensitivity of the minimal essential criteria once skin test positivity was replaced by *A.fumigatus*-specific IgE (criteria 3). Finally, criteria 7 based on immunological components alone had a reasonable performance on LCA but lower on the MDT evaluation due to the higher number of patients with bronchiectasis in the study.

We have previously used latent class analysis (LCA), a relatively modern computational approach for refining the diagnostic criteria for ABPA (Agarwal et al., 2013; Saxena et al., 2021). The LCA statistically calculates the relationship between certain user-defined input variables (for example, the various diagnostic criteria) that estimate a latent variable, namely the presence or absence of disease (ABPA) (Rindskopf and Rindskopf, 1986; Goetghebeur et al., 2000). Importantly, the LCA technique provides diagnostic accuracy measures without requiring a 2 x 2 matrix that is usually required when a reference standard is used. Conventionally, for performing LCA, we divide the asthma population into two groups. This procedure is complicated as

TABLE 4 | Performance of various diagnostic criteria for allergic bronchopulmonary aspergillosis (ABPA) using the multidisciplinary team (MDT) evaluation as the reference standard.

Criteria	MDT, Not ABPA (n=437)		MDT, ABPA (n=106)		Sensitivity (95% CI)	Specificity (95% Cl)	Correct classification rate (95% CI)	False-negative rate (95% CI)
	Present	Absent	Present	Absent	(,	(,		
Modified ISHAM-AWG criteria	0	437	102	4	96.2 (90.6-98.9)	100 (99.2-100)	99.3 (98.1-99.8)	3.8 (1.0-9.4)
New criteria								
Criteria 1	14	423	102	4	96.2 (90.6-98.9)	96.8 (94.7-98.2)	96.7 (94.8-98.0)	3.8 (1.0-9.4)
Criteria 2	24	413	102	4	96.2 (90.6-98.9)	94.5 (91.9-96.5)	94.8 (92.6-96.6)	3.8 (1.0-9.4)
Criteria 3	05	432	94	12	88.7 (81.1-94.0)	98.9 (97.4-99.6)	96.9 (95.0-98.2)	11.3 (5.9-18.9)
Criteria 4	02	435	78	28	73.6 (64.1-81.7)	99.5 (98.4-99.9)	94.5 (92.2-96.2)	26.5 (18.3-35.9)
Criteria 5	15	422	102	4	96.2 (90.6-98.9)	96.6 (94.4-98.1)	96.5 (94.6-97.9)	3.8 (1.0-9.4)
Criteria 6	31	406	102	4	96.2 (90.6-98.9)	92.9 (90.1-95.1)	93.6 (91.2-95.5)	3.8 (1.0-9.4)
Criteria 7	36	401	101	5	73.7 (65.5-80.9)	98.8 (97.2-99.6)	92.5 (89.9-64.5)	26.3 (19.1-34.4)

CI, confidence intervals; ISHAM-AWG, international society for human and animal mycology ABPA working group criteria.

The details of the various criteria are provided in Table 1.

each criterion (n=8) has to be applied in every subject. Further, the presence or absence of the criteria is reported in a binary fashion (0 or 1), and we have 256 different permutations in two groups (**Table S1**). We evaluated whether the results would be different if the asthma population is not divided into two groups (**Table S2**). We found the results similar whether or not the population is divided into two groups (**Table 3**).

What is the clinical relevance of our study? We evaluated seven simpler models for diagnosing ABPA. The A.fumigatusspecific IgE-based criteria performed better than skin test-based criteria. Of the seven, the combination of IgE (total and A.fumigatus-specific) and either an elevated A.fumigatusspecific IgG or bronchiectasis (criteria 5) could be a reasonable alternative to the modified ISHAM-AWG criteria. Also, the specific IgE-based minimal essential criteria (criteria 3) might be another alternative, especially in resource-constrained settings, as it includes only three components. The skin testbased minimal essential criteria (criteria 4) may be used to confirm ABPA (specificity >99%) in settings without access to immunoassays. However, criteria 4 cannot be used to rule out ABPA, given its poor sensitivity (73%). Lastly, our study results provide clinicians with evidence regarding the certainty of diagnosing ABPA when one or more of the components of the existing criterion are missing.

Finally, our study has a few limitations. Our study is a singlecenter study conducted at a tertiary care hospital. Also, a large proportion of patients in our study had bronchiectasis. The diagnostic performance of the criteria might be different in the milder asthmatics seen in the community or those with serological ABPA. Another weakness can be attributed to the statistical technique as the probabilistic estimates cannot be perfect and depend on the positivity and negativity rates of the variables included in the model. Also, the goodness-of-fit for the model indicated that conditional independence was not met. Thus, the diagnostic performance may be fallible. While an MDT comparison of the various diagnostic criteria was also performed to circumvent this flaw, the criteria need to be validated prospectively. The study results are also not immediately applicable to other predisposing conditions for ABPA, including cystic fibrosis and others.

In conclusion, we found that specific IgE-based simpler criteria may be good alternatives for diagnosing ABPA.

REFERENCES

- Agarwal, R., Aggarwal, A. N., Sehgal, I. S., Dhooria, S., Behera, D., and Chakrabarti, A. (2016). Utility of IgE (Total and Aspergillus Fumigatus Specific) in Monitoring for Response and Exacerbations in Allergic Bronchopulmonary Aspergillosis. *Mycoses* 59 (1), 1–6. doi: 10.1111/myc.12423
- Agarwal, R., Chakrabarti, A., Shah, A., Gupta, D., Meis, J. F., Guleria, R., et al. (2013). Allergic Bronchopulmonary Aspergillosis: Review of Literature and Proposal of New Diagnostic and Classification Criteria. *Clin. Exp. Allergy* 43 (8), 850–873. doi: 10.1111/cea.12141
- Agarwal, R., Dua, D., Choudhary, H., Aggarwal, A. N., Sehgal, I. S., Dhooria, S., et al. (2017). Role of Aspergillus Fumigatus-Specific IgG in Diagnosis and Monitoring Treatment Response in Allergic Bronchopulmonary Aspergillosis. *Mycoses* 60 (1), 33–39. doi: 10.1111/myc.12541

Because of the monocentric nature of our study, a prospective multicenter investigation is required for validating our study results.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Institute Ethics Committe, PGIMER, Chandigarh. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

RA conceived the idea, involved in patient management, statistical analysis, drafted, and revised the manuscript for intellectual content. PS: involved in patient management and data collection, revised the manuscript for intellectual content. VM: involved in patient management, statistical analysis, revised the manuscript. IS: involved in patient management, revised the manuscript. SD: involved in patient management, revised the manuscript. KP: involved in patient management, revised the manuscript for intellectual content. AA: involved in patient management, revised the manuscript for intellectual content. AC: involved in patient management, revised the manuscript for intellectual content. RA: guarantor of the paper. All authors contributed to the article and approved the submitted version.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fcimb.2022. 861866/full#supplementary-material

- Agarwal, R., Khan, A., Garg, M., Aggarwal, A. N., and Gupta, D. (2011). Pictorial Essay: Allergic Bronchopulmonary Aspergillosis. *Indian J. Radiol. Imaging* 21 (4), 242–252. doi: 10.4103/0971-3026.90680
- Agarwal, R., Khan, A., Gupta, D., Aggarwal, A. N., Saxena, A. K., and Chakrabarti, A. (2010). An Alternate Method of Classifying Allergic Bronchopulmonary Aspergillosis Based on High-Attenuation Mucus. *PloS One* 5 (12), e15346. doi: 10.1371/journal.pone.0015346
- Agarwal, R., Maskey, D., Aggarwal, A. N., Saikia, B., Garg, M., Gupta, D., et al. (2013). Diagnostic Performance of Various Tests and Criteria Employed in Allergic Bronchopulmonary Aspergillosis: A Latent Class Analysis. *PloS One* 8 (4), e61105. doi: 10.1371/journal.pone.0061105
- Agarwal, R., Muthu, V., Sehgal, I. S., Dhooria, S., Prasad, K. T., and Aggarwal, A. N. (2022). Allergic Bronchopulmonary Aspergillosis. *Clin. Chest Med.* 43 (1), 99–125. doi: 10.1016/j.ccm.2021.12.002

- Agarwal, R., Sehgal, I. S., Dhooria, S., and Aggarwal, A. N. (2016). Developments in the Diagnosis and Treatment of Allergic Bronchopulmonary Aspergillosis. *Expert Rev. Respir. Med.* 10 (12), 1317–1334. doi: 10.1080/17476348.2016. 1249853
- Aggarwal, A. N., Agarwal, R., Dhooria, S., Prasad, K. T., Sehgal, I. S., Muthu, V., et al. (2019). Joint Indian Chest Society-National College of Chest Physicians (India) Guidelines for Spirometry. *Lung India* 36 (Supplement), S1–S35. doi: 10.4103/lungindia.lungindia_300_18
- Asano, K., Hebisawa, A., Ishiguro, T., Takayanagi, N., Nakamura, Y., Suzuki, J., et al. (2021). New Clinical Diagnostic Criteria for Allergic Bronchopulmonary Aspergillosis/Mycosis and its Validation. J. Allergy Clin. Immunol. 147 (4), 1261–1268.e5. doi: 10.1016/j.jaci.2020.08.029
- Dhooria, S., and Agarwal, R. (2014). Diagnosis of Allergic Bronchopulmonary Aspergillosis: A Case-Based Approach. *Future Microbiol.* 9 (10), 1195–1208. doi: 10.2217/fmb.14.74
- Global Initiative for Asthma. (GINA). (2004). Global Strategy for Asthma Management and Prevention. Available at: https://ginasthma.org/wp-content/ uploads/2019/01/2004-GINA.pdf.
- Goetghebeur, E., Liinev, J., Boelaert, M., and van der Stuyft, P. (2000). Diagnostic Test Analyses in Search of Their Gold Standard: Latent Class Analyses With Random Effects. *Stat. Methods Med. Res.* 9 (3), 231–248. doi: 10.1177/ 096228020000900304
- Kosmidis, C., and Denning, D. W. (2015). The Clinical Spectrum of Pulmonary Aspergillosis. *Thorax* 70 (3), 270–277. doi: 10.1136/thoraxjnl-2014-206291
- Mackinnon, A. (2000). A Spreadsheet for the Calculation of Comprehensive Statistics for the Assessment of Diagnostic Tests and Inter-Rater Agreement. *Comput. Biol. Med.* 30 (3), 127–134. doi: 10.1016/S0010-4825(00)00006-8
- Pouillot, R., Gerbier, G., and Gardner, I. A. (2002). "TAGS", a Program for the Evaluation of Test Accuracy in the Absence of a Gold Standard. *Prev. Vet. Med.* 53 (1-2), 67–81. doi: 10.1016/S0167-5877(01)00272-0
- Rindskopf, D., and Rindskopf, W. (1986). The Value of Latent Class Analysis in Medical Diagnosis. *Stat. Med.* 5 (1), 21–27. doi: 10.1002/sim.4780050105

- Rosenberg, M., Patterson, R., Mintzer, R., Cooper, B. J., Roberts, M., and Harris, K. E. (1977). Clinical and Immunologic Criteria for the Diagnosis of Allergic Bronchopulmonary Aspergillosis. *Ann. Intern. Med.* 86 (4), 405–414. doi: 10.7326/0003-4819-86-4-405
- Saxena, P., Choudhary, H., Muthu, V., Sehgal, I. S., Dhooria, S., Prasad, K. T., et al. (2021). Which Are the Optimal Criteria for the Diagnosis of Allergic Bronchopulmonary Aspergillosis? A Latent Class Analysis. J. Allergy Clin. Immunol. Pract. 9 (1), 328–335.e1. doi: 10.1016/j.jaip.2020.08.043
- Schwartz, H. J., and Greenberger, P. A. (1991). The Prevalence of Allergic Bronchopulmonary Aspergillosis in Patients With Asthma, Determined by Serologic and Radiologic Criteria in Patients at Risk. J. Lab. Clin. Med. 117 (2), 138–142.

Conflict of Interest: RA has received grant support from Cipla, India on research in ABPA PS.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Agarwal, Saxena, Muthu, Sehgal, Dhooria, Prasad, Aggarwal and Chakrabarti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.